useful in confirming the configuration of the PGB compounds at C₁₅ and for estimating the composition of mixtures of enantiomers, 9.10 since the (+)- α -methoxy- α -trifluoromethylphenyl acetates (MTPA) of (15R)- and (15S)-PGB₂ methyl esters showed several differences in the proton and fluorine nuclear magnetic resonance spectra.¹¹

The prostaglandins extracted from a specimen of the S form of P. homomalla after enzymatic hydrolysis were purified by column chromatography. Based on frozen wet weight of coral, the amounts of (15S)-PGA₂ (1.4%) and (15S)-PGA₂ methyl ester (0.4%) obtained are comparable to the amounts of (15R)-prostaglandins obtained from the (R) form of P. homomalla. In addition,⁶ 0.06% of crystalline (15S)-PGE₂, mp 63-66.5°, was isolated and shown to be identical in physical and biological properties with mammalian PGE2.12

These findings, in addition to providing a novel and possibly useful natural source of primary prostaglandins,6 also raise many intriguing biochemical questions about the origin and role of prostaglandins in marine organisms, some of which are under investigation.

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(11) In the proton magnetic spectrum, the doublet from the proton at C₁₃ occurred centered at δ 6.79 (J = 16 Hz) for the (15S) isomer and at 6.89 (J = 16 Hz) for the (15R) isomer. The chemical-shift differences for the methoxyl protons were too small to be useful since they were closely coupled quartets due to splitting by the three fluorine atoms five bonds distant (Varian A-60-A, CDCl₈, tetramethylsilane internal reference). The fluorine magnetic resonance spectrum (obtained from Midwest Research Institute at 94.1 MHz in CDCl₃ relative to external trifluoroacetic acid) of the 15-epimers showed absorption frequencies of 675((R) isomer) and 687((S) isomer) for the trifluoromethyl groups, broadened enough by splitting from the methoxyl protons to make accurate integration difficult without comparison with computer-calculated spectra.

(12) Infrared and nuclear magnetic resonance spectroscopy, mixture melting point, and chromatographic behavior (several systems) all were identical. We are grateful to Dr. J. R. Weeks and coworkers, The Upjohn Company, for the biological assays.

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The Synthesis of Prostaglandins E_2 and $F_{2\alpha}$ from (15R)- and (15S)-PGA₂

Sir:

Ester derivatives of both (15S)- and (15R)-PGA₂ are readily obtainable from the gorgonian Plexaura homomalla found in the Caribbean area.^{1,2} This communication describes the conversion of these materials to the biologically important³ primary prostaglandins PGE_2 and $PGF_{2\alpha}$. (15S)-Prostaglandin A₂, acetate, methyl ester (1a) from coral extracts² was epoxidized with alkaline hydrogen peroxide⁴ to a mixture of isomeric 10,11-epoxides (2a). Without sep-



aration, the mixture was reduced with chromous acetate⁵ in acetic acid or aluminum amalgam⁶ to give, after separation by silica gel chromatography, 3a, the 15acetate, methyl ester of PGE₂, in 56% yield along with 25% of the corresponding 11-epimer. The diester 3a was hydrolyzed⁷ to give PGE₂ (4a), obtained crystalline, mp 66-68°, in 90% yield, and identical in all respects⁸ with PGE₂ obtained from mammalian sources. The 11β isomer of PGE₂ obtained similarly was noncrystalline, slightly less polar than 4a on silica gel, showing characteristic downfield shifts of the C13,14 olefinic protons in the nmr spectrum and characteristic fine structure differences in the circular dichroism curve.9

Reduction of PGE₂ with sodium borohydride leads directly to $PGF_{2\alpha}$.¹⁰ Compound **3a** was also converted to its 11-trimethylsilyl ether and reduced with sodium borohydride to a mixture of 9α and 9β alcohols separated by silica gel chromatography; use of this protecting group increases the 9α : 9β ratio obtained on reduction of the 9-ketone.¹¹ After hydrolysis $PGF_{2\alpha}$

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(6a), mp 25-35°, was obtained from 3a in 60% yield and was converted to its tris(hydroxymethyl)aminomethane (THAM) salt,¹² mp 100-101°. Both PGF_{2α} and its THAM salt were identical with authentic materials.

Utilization of the (15R)-PGA₂ diester (1b) from coral as a precursor of PGE₂ and PGF_{2 α} requires an inversion of configuration at C-15.¹¹ For the synthesis of PGF_{2 α}, 1b was carried through the same sequence as above giving the corresponding intermediates 2b, 3b, and 5b. On hydrolysis 5b gave 6b, the 15-epimer of PGF_{2 α}.

Selective oxidation with 2,3-dichloro-5,6-dicyano-1,4-benzoquinone¹³ gave ketone 7 (λ_{max} 234 nm (ϵ 11,850)) which was reduced by zinc borohydride in dimethoxyethane¹⁴ after temporary protection of the hydroxyl groups by trimethylsilylation, giving a 73:27 ratio of PGF_{2α} (**6a**) and its 15-epimer **6b**.

(15*R*)-PGA₂ methyl ester (8b), also available from coral, was treated with methanesulfonyl chloride in pyridine and the resulting crude 15-mesylate was solvolyzed in acetone-water to give modest yields of the C_{15} inverted product, (15*S*)-PGA₂ methyl ester (8a), along with some 8b and several other products. Acetylation of 8a in acetic anhydride-pyridine gave 1a and thus ultimately PGE₂ and PGF_{2α}.

Plexaura homomalla, var. (R) and var. (S), are thus both suitable sources of (coral) prostaglandins useful in the synthesis of PGE₂ and PGF_{2 α}. From the (S) variety, PGE₂ can be obtained in three steps and PGF_{2 α} in four steps.

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Isolation of a New Naturally Occurring Prostaglandin, 5-trans-PGA₂. Synthesis of 5-trans-PGE₂ and 5-trans-PGF_{2 α}

Sir:

During the chromatographic purification of (15S)-PGA₂ obtained from the gorgonian *Plexaura homomalla* var. (S),¹ a new natural prostaglandin was detected which was chromatographically less polar than PGA₂ on silver nitrate impregnated silica gel. We report here the purification of this material, its structure elucidation, and confirmation of the structure by chemical transformations.

Column chromatography of crude (15S)-PGA₂ on Amberlyst-15 Ag⁺ form² or on silver nitrate impregnated silica gel gave a minor component to which the structure (15S)-15-hydroxy-9-oxo-5-trans,10,13-transprostatrienoic acid (5-trans-PGA₂) (1) is assigned. Content of the trans isomer usually ranged between 5 and 15% of the PGA₂ present. 5-trans-PGA₂ is an oil [λ_{max} 217 nm (ϵ 9050); [α]D +128° (CHCl₃); molecular ion at 478.2998 for TMS derivative (calcd for

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C₂₆H₄₆O₄Si₂, 478.2932); mass spectrum identical with that of PGA₂]. Conversion of **1** to the β -ketol was effected by a modification of the epoxidation-reduction sequence³ to give (15S)-11 α ,15-dihydroxy-9-oxo-5trans,13-trans-prostadienoic acid (5-trans-PGE₂) (2)⁴ together with the 11 β isomer. 5-trans-PGE₂ was crystalline: mp 76-77° (Anal. Found: C, 68.52; H, 9.23); [α]D -66° (c 0.983, ethanol); mass spectrum identical with PGE₂. After conversion to a trimethylsilyl (TMS) derivative, reduction of 2 with sodium borohydride⁵ and hydrolysis gave a mixture of (15S)-9 α ,11 α ,15-trihydroxy-5-trans,13-trans-prostadienoic acid (5-trans-PGF_{2 α}) (3) and (15S)-9 β ,11 α ,15-trihydroxy-5trans,13-trans-prostadienoic acid (5-trans-PGF_{2 β}) (4),



which were separated by silica gel chromatography. 5-trans-PGF_{2 α} was crystalline: mp 94.8–95.8° (Anal. Found: C, 67.99; H, 9.64); [α]D +9° (ethanol); mass spectrum m/e at 354 (M⁺), 336, 318, 264, 247, 191, 137. 5-trans-PGF_{2 β} was also crystalline: mp 68–69° (Anal. Found: C, 67.89; H, 9.78); [α]D -8° (ethanol).

Irradiation of prostaglandin E_2 in oxygen-free benzene-methanol solution with 3500-Å light for 24 hr in a Rayonet photochemical reactor in the presence of diphenyl sulfide^{6,7} gave, after careful chromatography on acid-washed silica gel, a 22 % yield of 5-*trans*-PGE₂, mp 75-77°, which was identical with the material derived from *P. homomalla*. In a similar fashion and in similar yield, crystalline 5-*trans*-PGF_{2β} and 5-*trans*-PGF_{2α} were prepared from the corresponding 5-*cis*prostaglandins and were also identical with the coralderived compounds.

A reexamination of the extracts of *P. homomalla* var. (S) prior to hydrolysis shows that, while small amounts of the free acids are present, the 5-trans isomer is predominantly in the form of its 15-acetate methyl ester. It is not clear at this time whether the presence of this isomer represents biosynthetic formation from 5-trans-arachidonic acid endogenous to *P. homomalla*, or a subsequent transformation product of 5-cis-PGA₂.

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